

Yeast Screen Assay Card

Hybridization Probe Assays

This reagent kit is designed to test for DNA from *Yeast (S. cerevisiae)* from two unknown liquid or dry samples. These procedures are to be performed manually and have been simplified to reduce the risk of operator error.

⚠ WARNING: If the unknown sample to be tested is suspected to contain a pathogen or toxin, it must be handled by trained personnel and treated as if it were highly poisonous or infectious throughout the sample purification procedure, the assay procedure, and when disposing of all waste materials that have come in contact with the suspected sample.

Kit Contents

Kit Part No. PATH-ASY-0006

5 mL Reagent Grade Water (1)	5 mL Unknown Sample Bottles (2)	1 mL Syringes with Cannula Tips (4)
Sample Swab Pack (1)	Instruction Booklet (1)	Freeze-dried Reagents in a Foil Bag (1)
Transfer Pipets Pack (1)	Loading Instructions Sticker (1)	

RAZOR® (Part # RAZR-ASY-0003, RAZR-ASY-0013) or RAZOR® EX (RAZR-RED-4000) is required

Programming the Run Protocol

Before the sample is loaded into the pouch, verify that the run protocol for this test is loaded onto the instrument. If it is not and you are using a RAZOR, use the RAZOR desktop software to create the protocol and load it onto the instrument. Use the data in this card when creating the protocol for this particular test with the Protocol Wizard. For the RAZOR EX, you can load the protocol by scanning the square protocol bar code on the reagent box. If the bar code is damaged, use the generic bar codes that are located below. Times, temperatures, and results analyses are all included in the protocol bar code.

Loading a Protocol for the RAZOR® EX

After the square protocol bar code has been scanned, scan the rectangular bar code on the pouch fitment. If the rectangular bar code is damaged, use the generic one below. **Note: You will only be able to use the generic bar code for one run. In order to re-use it, you have to delete the run.**



ITYEASTB-ITYST001



Kit Part Number: PATH-ASY-0006
Protocol Code: ITYEASTB

Step 1:

Negative Port

Add 0.5 mL reagent grade water.

Step 2:

Unknown 1 Port

Add 0.5 mL sample 1.

Step 3:

Unknown 2 Port

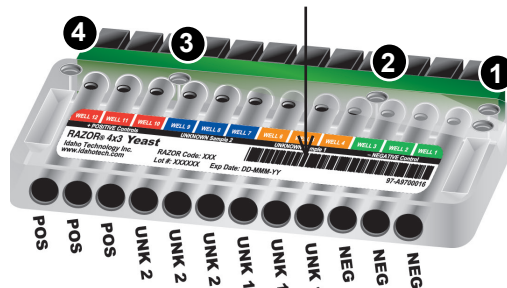
Add 0.5 mL sample 2.

Step 4:


Positive Port

Add 0.5 mL reagent grade water.

Rectangular Bar Code



Protocol Steps for the RAZOR® EX

Protocol Steps	Data to be Entered																																																																																				
Step 1: Protocol Identifier	Enter: 06H to identify this protocol. This identifying code matches the first three characters on the pouch. This will help you identify the protocol after it is loaded on the instrument.																																																																																				
Step 2: Protocol Parameters	Initial Hold Temperature: 93°C Denature Temperature: 91°C Anneal Temperature: 60°C																																																																																				
	Initial Hold Time: 120 s Denature Hold Time: 3 s Anneal Hold Time: 15 s																																																																																				
	Cycles: 55																																																																																				
Step 3: Select Organisms for the Protocol	A list of organisms is listed in the All Organisms column. Select Yeast and click the Add button. Yeast has just been added to the Organisms for this Protocol column. If Yeast is not listed, see the <i>Pouch Instruction Booklet</i> for instructions on how to add new organisms. Note: The target for Yeast is "Tar1."																																																																																				
Step 4: Add Environmental Samples	Add two unknown samples by clicking the Add button and entering UNK1 . Repeat the step and enter UNK2 .																																																																																				
Step 5: Assign an Organism to Each Sample Position	This step should not appear for this protocol. If it does appear, you have selected multiple organisms in step 3. Be sure that only one organism is selected in step 3.																																																																																				
Step 6: RAZOR Pouch Definition	<p>Fill out the definition as shown in the image below</p> <table border="1"> <thead> <tr> <th colspan="6">Razor Bag Definition</th> </tr> <tr> <th>Pos</th> <th>Organism</th> <th>Target</th> <th>Type</th> <th>Env. Sample</th> <th>Concentration</th> </tr> </thead> <tbody> <tr><td>1</td><td>Yeast</td><td>Tar1</td><td>NEG</td><td></td><td></td></tr> <tr><td>2</td><td>Yeast</td><td>Tar1</td><td>NEG</td><td></td><td></td></tr> <tr><td>3</td><td>Yeast</td><td>Tar1</td><td>NEG</td><td></td><td></td></tr> <tr><td>4</td><td>Yeast</td><td>Tar1</td><td>UNK</td><td>UNK1</td><td></td></tr> <tr><td>5</td><td>Yeast</td><td>Tar1</td><td>UNK</td><td>UNK1</td><td></td></tr> <tr><td>6</td><td>Yeast</td><td>Tar1</td><td>UNK</td><td>UNK1</td><td></td></tr> <tr><td>7</td><td>Yeast</td><td>Tar1</td><td>UNK</td><td>UNK2</td><td></td></tr> <tr><td>8</td><td>Yeast</td><td>Tar1</td><td>UNK</td><td>UNK2</td><td></td></tr> <tr><td>9</td><td>Yeast</td><td>Tar1</td><td>UNK</td><td>UNK2</td><td></td></tr> <tr><td>10</td><td>PCR POS</td><td>Tar1</td><td>POS</td><td></td><td></td></tr> <tr><td>11</td><td>PCR POS</td><td>Tar1</td><td>POS</td><td></td><td></td></tr> <tr><td>12</td><td>PCR POS</td><td>Tar1</td><td>POS</td><td></td><td></td></tr> </tbody> </table>	Razor Bag Definition						Pos	Organism	Target	Type	Env. Sample	Concentration	1	Yeast	Tar1	NEG			2	Yeast	Tar1	NEG			3	Yeast	Tar1	NEG			4	Yeast	Tar1	UNK	UNK1		5	Yeast	Tar1	UNK	UNK1		6	Yeast	Tar1	UNK	UNK1		7	Yeast	Tar1	UNK	UNK2		8	Yeast	Tar1	UNK	UNK2		9	Yeast	Tar1	UNK	UNK2		10	PCR POS	Tar1	POS			11	PCR POS	Tar1	POS			12	PCR POS	Tar1	POS		
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Step 7: RAZOR Metacalls	This step displays an image of the conditions required for an unknown to be called positive. You do not need to enter any data in this step. Click Finish to save the protocol.																																																																																				
NOTE:	<p>Before interpreting results on machine's LED screen, remember assays correlate to the channels shown in diagram to right.</p>  <p>The diagram shows 12 assay slots labeled Slot 1 through Slot 12. Below the slots is a green LED screen displaying the text: "Call:+++ --- +++ ---" and "NEXT PREV" on the left, and "DONE" on the right.</p>																																																																																				